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The LBFGS quasi-Newtonian method for molecular modeling prion AGAAAAGA amyloid fibrils

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ABSTRACT

Experimental X-ray crystallography, NMR (Nuclear Magnetic Resonance) spectroscopy, dual polarization interferometry, etc. are indeed very powerful tools to determine the 3-Dimensional structure of a protein (including the membrane protein); theoretical mathematical and physical computational approaches can also allow us to obtain a description of the protein 3D structure at a submicroscopic level for some unstable, noncrystalline and insoluble proteins. X-ray crystallography finds the X-ray final structure of a protein, which usually need refinements using theoretical protocols in order to produce a better structure. This means theoretical methods are also important in determinations of protein structures. Optimization is always needed in the computer-aided drug design, structure-based drug design, molecular dynamics, and quantum and molecular mechanics. This paper introduces some optimization algorithms used in these research fields and presents a new theoretical computational method—An improved LBFGS Quasi-Newtonian mathematical optimization method—to produce 3D structures of prion AGAAAAGA amyloid fibrils (which are unstable, noncrystalline and insoluble), from the potential energy minimization point of view. Because the NMR or X-ray structure of the hydrophobic region AGAAAAGA of prion proteins has not yet been determined, the model constructed by this paper can be used as a reference for experimental studies on this region, and may be useful in furthering the goals of medicinal chemistry in this field.

Keywords: Protein 3D Structure; Computational

Approaches; Optimization Method; Molecular Modelling; Prion AGAAAAGA Amyloid Fibrils

1. INTRODUCTION

Neurodegenerative diseases including Parkinson's, Alzheimer's, Huntington's, and Prion's were found they all featured amyloid fibrils [1-6]. Amyloid is characterized by a cross- β sheet quaternary structure and recent X-ray diffraction studies of microcrystals revealed atomistic details of core region of amyloid [7,8]. All the quaternary structures of amyloid cross- β spines can be reduced to one of the 8 classes of steric zippers of [8], with strong van der Waals (vdW) interactions between β -sheets and hydrogen bonds (HBs) to maintain the β -strands. A new era in the structural analysis of amyloids started from the “steric zipper”- β -sheets [7]. As the two β -sheets zip up, Hydrophobic Packings (HPs & vdWs) have been formed. The extension of the “steric zipper” above and below (*i.e.* the β -strands) is maintained by Hydrogen Bonds (HBs) (but usually there is no HB between the two β -sheets). This is the common structure associated with some 20 neurodegenerative amyloid diseases.

We first do some mathematical analysis for the common structure. The vdW contacts of the two atoms can be described by the Lennard-Jones (LJ) potential energy, where the parameters on the depth of the potential well and the atom diameter can be fitted to reproduce experimental data or deduced from results of accurate quantum chemistry calculations. There are two parts describing LJ potential energy: the repulsion and the attraction (**Figure 1**). The minimization of LJ potential energy is an optimization problem, which is a well-known and challenging test problem for global optimization [10-14]. It is very hard for global optimization to directly solve this problem even with a small number of atoms. Similarly,

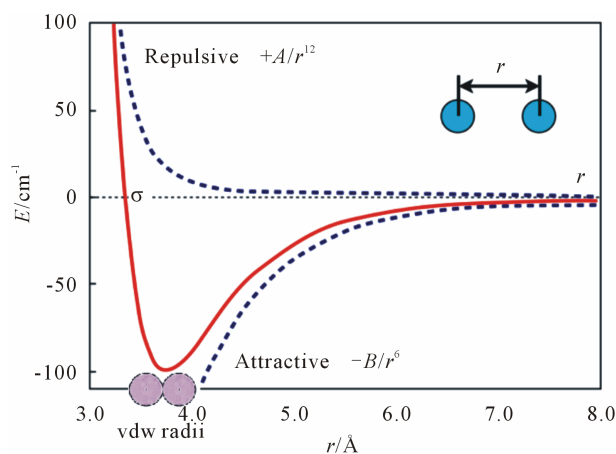


Figure 1. The LJ potential energy (see **Figure 1** of [9]).

the potential energy for the HBs between β -strands has a similar mathematical formula, and usually most of the HBs are still kept during the phase of molecular modelling of amyloid fibrils. Thus, the amyloid fibril molecular modelling problem can be reduced to solve the optimization problem of minimizing LJ potential energy though it is not easy to accurately solve for a large molecule.

Alternatively, we have found another way to solve the problem of minimizing LJ potential energy [15]. Seeing **Figure 1**, we may know that this optimization problem reaches its optimal value at the bottom of the LJ potential well, where the distance between two atoms equals to the sum of vdW radii of the two atoms. Hence, the amyloid fibril molecular modelling problem can be looked as a molecular distance geometry problem (MDGP) [16-18]. As an example to explain MDGP, the problem of locating sensors in telecommunication networks is a DGP. In such a case, the positions of some sensors are known (which are called anchors) and some of the distances between sensors (which can or cannot be anchors) are known. The DGP is to locate the positions of all the sensors. The MDGP looks sensors as atoms and their telecommunication network as a molecule. In this paper, we aim to solve the alternative MDGP problem (*i.e.* **Eq.11** of [15]) for modelling amyloid fibril molecular 3D structures.

Neurodegeneration is the progressive loss of structure or function of neurons, including death of neurons. A prion is a misshapen protein that acts like an infectious agent (but not requiring either DNA, RNA, or both) to cause a number of fatal diseases. Prion diseases are rich in β -sheets (compared with the normal prion protein PrPC in rich of α -helices) and are so-called “protein structural conformational” diseases. The normal hydrophobic region 113-120 AGAAAAGA peptide of prion proteins is an inhibitor/blocker of prion diseases. PrP lacking this palindrome could not convert to prion diseases. Brown *et al.* pointed out that the AGAAAAGA

peptide was found to be necessary (though not sufficient) for blocking the toxicity and amyloidogenicity of PrP 106-126, and the peptide AGAA does not form fibrils [19]. The minimum sequence necessary for fibril formation should be AGAAA, AGAAAA, AGAAAAG, AGAAAAGA and GAAAAGA, but the molecular structures of these fibrils have not known yet. This paper addresses an important problem on modelling the 3D molecular structures of prion AGAAAAGA amyloid fibrils of neurodegenerative diseases. The rest of this paper is arranged as follows. In the next section, *i.e.* Section 2, an improved LBFGS Quasi-Newtonian method is presented for solving **Eq.11** of [5]. Section 3 implements this Quasi-Newtonian method by constructing a 3D molecular structure of prion AGAAAAGA amyloid fibrils of neurodegenerative prion diseases. Numerical results of computations show that the method designed in Section 2 is very effective and successful. This concluding remark will be made in the last section, *i.e.* Section 4.

2. METHODS

In a (macro) molecular system, if it is very far from equilibrium, then the forces may be excessively large, a robust energy minimization (EM) is required; another reason to perform an EM is the removal of all kinetic energy from the system: EM reduces the thermal noise in the structures and potential energies [20]. EM, with the images at the endpoints fixed in space, of the total system energy provides a minimum energy path. EM can be done using steepest descent (SD), conjugate gradient (CG), and Limited-memory Broyden Fletcher Goldfarb Shanno (LBFGS) methods.

Three kinds of possible EM methods are: 1) Derivative-free methods—that require only function evaluations, *e.g.* the simplex method and its variants; 2) derivative information methods—the partial derivatives of the potential energy with respect to all coordinates are known and the forces are minimized, *e.g.* SD, CG methods; and 3) second derivative information methods, *e.g.* LBFGS method. “SD is based on the observation that if the real-valued function $f(x)$ is defined and differentiable in a neighbourhood of a point x_0 then $f(x)$ decreases fastest if one goes from x_0 in the direction of the negative gradient of $f(x)$ at x_0 and SD local search method converges fast [21]. SD is robust and easy to implement but it is not most efficient especially when closer to minimum; at this moment, we may use the efficient CG. CG is slower than SD in the early stages but more efficient when closer to minimum. CG algorithm adds an orthogonal vector to the current direction of the search, and then moves them in another direction nearly perpendicular to this vector. The hybrid of SD-CG will make SD or CG more efficient than SD or CG alone. However,

CG cannot be used to find the EM path, for example, when “forces are truncated according to the tangent direction, making it impossible to define a Lagrangian” [22,23]. In this case, the powerful and faster quasi-Newtonian method (e.g. the LBFGS quasi-Newtonian minimiser) can be used [22,24-28]. We briefly introduce the LBFGS quasi-Newtonian method as follows.

Newton’s method in optimization explicitly calculates the Hessian matrix of the second-order derivatives of the objective function and the reverse of the Hessian matrix [29]. The convergence of this method is quadratic, so it is faster than SD or CG. In high dimensions, finding the inverse of the Hessian is very expensive. In some cases, the Hessian is a non-invertible matrix, and furthermore in some cases, the Hessian is symmetric indefinite. Quasi-Newton methods thus appear to overcome all these shortcomings.

Quasi-Newton methods (a special case of variable metric methods) are to approximate the Hessian. Currently, the most common quasi-Newton algorithms are the SR1 formula, the BHHH method, the widespread BFGS method and its limited/low-memory extension LBFGS, DFP, MS, and Broyden’s methods [30-33]. In Amber [23] and Gromacs [20], LBFGS is used, and the hybrid of LBFGS with CG—a Truncated Newton linear CG method with optional LBFGS Preconditioning [25]—is used in Amber [23].

For BFGS method, whether it converges at all on non-convex problems is still an open problem. In fact, Powell (1984) gave a counter-example that shows that BFGS with an inexact line-search search may fail to converge [34-36]. Li and Fukushima (2001) proposed a modified BFGS method for non-convex objective function [37]. Basing on [24,25,37-39], in this paper we present an improved LBFGS method described as follows [40]—which presents the non-monotone line search technique [41,42] for the Wolfe-type search. The improved LBFGS method presented in this paper is much better than the standard BFGS method in view of the CPU time (see **Figure 2**) tested through more than 30 nonlinear programming problems (where each selected problem is regular, that is, its first and second derivatives exist and are continuous everywhere, and each problem is with different dimensions, *i.e.*, 100, 500, 1000 and 10,000 dimensions) and its mathematical theory to support this algorithm can be seen from [40]. This paper implements the Wolfe-type search by the approximation technique of piecewise linear or quadratic function [43].

Algorithm 1: An Improved LBFGS Optimization Method for minimizing a non-convex function [40].

Step 0: Choose an initial point $x_0 \in R^n$, an initial positive definite matrix H_0 , and choose constants σ_1, σ_2 such that $0 < \sigma_1 < \sigma_2 < 1$, and choose an positive integer m_1 . Let $k=0$.

Step 1: If $\|g_k\| = 0$, then output x_k and stop; otherwise,

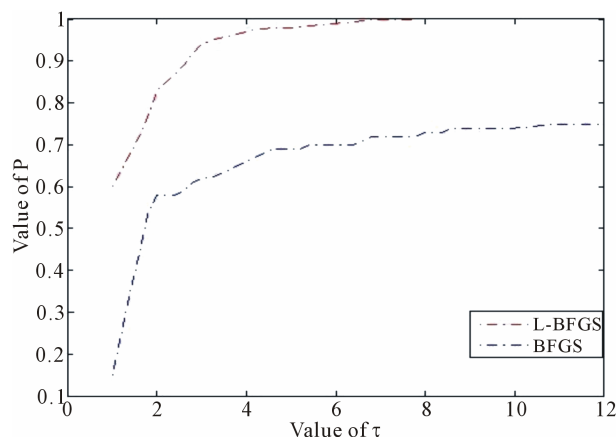


Figure 2. Performance based on CPU time, where $\tau \in R^1$ is a factor of the best possible ratio for CPU performance ratio of solvers L-BFGS and BFGS, and P is the probability distribution of the CPU performance ratio. Detailed mathematical formula for P can be seen in [44].

go to **Step 2**.

Step 2: Solve the following linear equation to get d_k :

$$d_k = -H_k g_k,$$

Step 3: Find a step-size $\lambda_k > 0$ satisfying the Wolfe-type line search conditions:

$$f(x_k + \lambda_k d_k) \leq f(x_k) + \sigma_1 \lambda_k g_k^T d_k,$$

$$g(x_k + \lambda_k d_k)^T d_k \geq \sigma_2 g_k^T d_k.$$

Step 4: Let $x_{k+1} = x_k + \lambda_k d_k$ be the next iteration. Calculate g_{k+1} and $\|g_{k+1}\|$.

Step 5: Let $s_k = x_{k+1} - x_k = \lambda_k d_k$, $y_k = g_{k+1} - g_k$, $\gamma_k = \|g_k\|$, then $y_k^* = y_k + \gamma_k s_k$.

Step 6: Let $m = \min\{k+1, m_1\}$. Update H_k following the formula

$$H_{k+1} = (V_k^{*T} \cdots V_{k-m+1}^{*T}) H_{k-m+1} (V_{k-m+1}^* \cdots V_k^*) \\ + \omega_{k-m+1}^* (V_{k-1}^{*T} \cdots V_{k-m+2}^{*T}) S_{k-m+1} S_{k-m+1}^T (V_{k-m+2}^* \cdots V_{k-1}^*) \\ + \cdots + \omega_k^* S_k S_k^T,$$

where $\omega_k^* = 1/(S_k^T y_k^*)$, $V_k^* = I - \omega_k^* y_k^* S_k^T$ (when $k=0$,

$$H_0 = y_0^T S_0 / (\|y_0\|^2), \quad V_0^* = I - y_0^* S_0^T / (S_0^T y_0^*),$$

$$H_1 = (I - y_0^* S_0^T / (S_0^T y_0^*))^T H_0 (I - y_0^* S_0^T / (S_0^T y_0^*)) \\ + 1/(S_0^T y_0^*) S_0 S_0^T.$$

Step 7: Let $k = k+1$ and go to **Step 1**.

In Algorithm 1, g_k denotes the gradient of $f(x)$ at x_k , and the convergence and the R-linear convergent rate of Algorithm 1 are guaranteed by the following assumptions: (A1) f is twice continuously differentiable, (A2) f has Lipschitz continuous gradients and Hessians, (A3) the

Hessian at the stationary point is always positively definite, and (A4) the level set of f is bounded. The detailed proof of the convergence and R-linear convergent rate of Algorithm 1 can be found in [40].

In [40], the non-monotone line search technique for step-size λ_k is used. This is a difference between the algorithm of [40] and this paper. All in all, it is well known that quasi-Newton method is an efficient solution method for unconstrained and continuously differentiable minimization problem [45-47]. However, it needs computing and storage of the updated matrix which is an approximation to the Hessian matrix in each iteration of the method. Hence, its efficiency may decrease when it is applied to large scale optimization problem. To overcome the drawback, limited memory quasi-Newton method is proposed [48]. The main ideal of this method is nearly identical to that of the standard BFGS method, and the only difference is that the inverse Hessian approximation is not formed explicitly, but defined by a small number, say m , of BFGS updates. This technique received much attention in recent years and numerical experiments show that it is very competitive [24,49], and its global convergence and R-linear convergence rate with Wolfe line search are established for the uniformly convex case [24,26]. Since the limited memory BFGS method may suffer from ill-conditions for small value of m , Al-Baali (1999) [50] made some modifications to the method and establish its global convergence based on the same assumptions, and Byrd *et al.* (1994) [51] derives new representation of limited memory quasi-Newton matrices for the benefit of computing the updated matrix. Recently, a non-monotone line search is introduced, see e.g., [41,42]. Then it is showed to be more competitive and practical for solving nonlinear optimization problems, and [52] established the global convergence of this line search applied to limited memory BFGS method based on the uniformly convex assumption. Motivated by the above observation, it turns out that in two respects the limited memory BFGS method is much less effective. First, we note that the convergence analysis of the method is focused on the uniformly convex assumption and little is known for non-convex case. Second, numerical experiments have suggested the main weakness of limited memory method is that it may converge very slowly in terms of number of iterations for ill-conditioned problems. The purpose of the above Algorithm 1 is to reduce these defects and **Figure 2** shows the effectiveness of the proposed algorithm. We will apply it into the molecular modelling of prion AGAAAAGA amyloid fibrils in the next section.

3. RESULTS AND DISCUSSION

From their research of prion, scientists found that the

cross- β structure of peptides is with the nature of self-aggregation, the self-aggregating to form fibers. This provides us a new research idea for nanomaterials. HBs can be formed between peptide β -strands, and one peptide monomer connects together with another in accordance with the specific structure to form fibers. Many laboratories in the world are synthesizing peptides that can self-aggregate to form fibers, and want to be able to control the growth of the fiber to find out new functional materials [53,54]. The studies of this paper not only benefit nanomaterials research, but also benefit the research on neurodegenerative amyloid fibril diseases. Prion AGAAAAGA peptide has been reported to own an amyloid fibril forming property (initially described in 1992 by Gasset *et al.* of Prusiner's Group) [19,55-78], but there has not been experimental structural bioinformatics for this segment yet due to the unstable, noncrystalline and insoluble nature of this region. Furthermore, Zhang (2011) did accurate calculations to confirm the amyloid fibril property at this region (**Figure 3**) [79] in the use of the fibril prediction program of [80].

3.1. Material for the Molecular Modeling

This paper uses a suitable pdb file template 2OMP.pdb (the LYQLEN peptide derived from human insulin residues 13-18 [8]) from the Protein Data Bank to build an 8-chain AGAAAAGA prion amyloid fibril molecular model to illuminate Algorithm 1 works very well. To choose 2OMP.pdb (**Figure 4**) as the modeling template is due to it can pass all the long procedures of SDCG-SA (equilibrations & productions)-SDCG of [79] in the use of the fibril prediction program of [80]. By observations of **Figure 4** and the 2nd column of coordinates of 2OMP.pdb, we know that E(F) chains can be calculated on the XZ-plane from A(B) chains by **Eq.3.1** and other chains can be got by a parallel up (or down) along the X-axis by **Eqs.3.2-3.3**:

$$E(F) = A(B) + (-1.885, 0, 17.243), \quad (3.1)$$

$$C(D) = A(B) + (9.666, 0, 0) \quad (3.2)$$

$$G(H) = E(F) + (9.666, 0, 0) \quad (3.3)$$

3.2. New Molecular Modeling Homology Model

Basing on the template 2OMP.pdb from the Protein Data Bank (**Figure 4**), the AGAAAAGA palindrome amyloid fibril model of prions (denoted as Model 1) will be constructed. Chains AB of Model 1 will be got from AB Chains of 2OMP.pdb using themutatemodule of the free package Swiss-PdbViewer (SPDBV Version 4.01). It is pleasant to see that some HBs are still kept after the mutations; thus we just need to consider the vdWs only. Making mutations for EF Chains of 2OMP.pdb, we can

get the EF Chains of Model 1. Then we add GLY and ALA residues by XLEaP module of Amber 11. However, the vdWs between Chain A and Chain E, between B Chain and F Chain are too far at this moment (**Figure 5**, where the twice of the vdW radius of CB atom is 3.4 angstroms).

In [79] the commercial package InsightII (accelrys.com)

is used to build models. Instead of InsightII, because this package is not available by the authors, this paper uses Algorithm 1 to build and optimize Model 1. In “Zipper 1”, fixing the coordinates of A.ALA3.CB, A.ALA1.CB and letting the coordinates of E.ALA6.CB, E.ALA4.CB be variables, we may get an optimization problem:

$$\begin{aligned}
 &\text{minimize} \\
 &4 \left\{ \frac{1}{\left[(x_{11}-1.071)^2 + (x_{12}-2.986)^2 + (x_{13}-1.888)^2 \right]^6} \right\} \\
 &-1 \left\{ \frac{1}{\left[(x_{11}-1.071)^2 + (x_{12}-2.986)^2 + (x_{13}-1.888)^2 \right]^3} \right\} \\
 &+4 \left\{ \frac{1}{\left[(x_{21}-1.071)^2 + (x_{22}-2.986)^2 + (x_{23}-1.888)^2 \right]^6} \right\} \\
 &-1 \left\{ \frac{1}{\left[(x_{21}-1.071)^2 + (x_{22}-2.986)^2 + (x_{23}-1.888)^2 \right]^3} \right\} \\
 &+4 \left\{ \frac{1}{\left[(x_{21}-1.135)^2 + (x_{22}+0.763)^2 + (x_{23}-7.209)^2 \right]^6} \right\} \\
 &-1 \left\{ \frac{1}{\left[(x_{21}-1.135)^2 + (x_{22}+0.763)^2 + (x_{23}-7.209)^2 \right]^3} \right\}
 \end{aligned} \tag{3.4}$$

or

$$\begin{aligned}
 &\text{minimize} \\
 &\frac{1}{2} \left\{ (x_{11}-1.071)^2 + (x_{12}-2.986)^2 + (x_{13}-1.888)^2 - 3.42^2 \right\}^2 \\
 &+ \frac{1}{2} \left\{ (x_{21}-1.071)^2 + (x_{22}-2.986)^2 + (x_{23}-1.888)^2 - 3.42^2 \right\}^2 \\
 &+ \frac{1}{2} \left\{ (x_{21}-1.135)^2 + (x_{22}+0.763)^2 + (x_{23}-7.209)^2 - 3.42^2 \right\}^2 \\
 &-0.05(x_{11} + x_{12} + x_{13} + x_{21} + x_{22} + x_{23})
 \end{aligned} \tag{3.5}$$

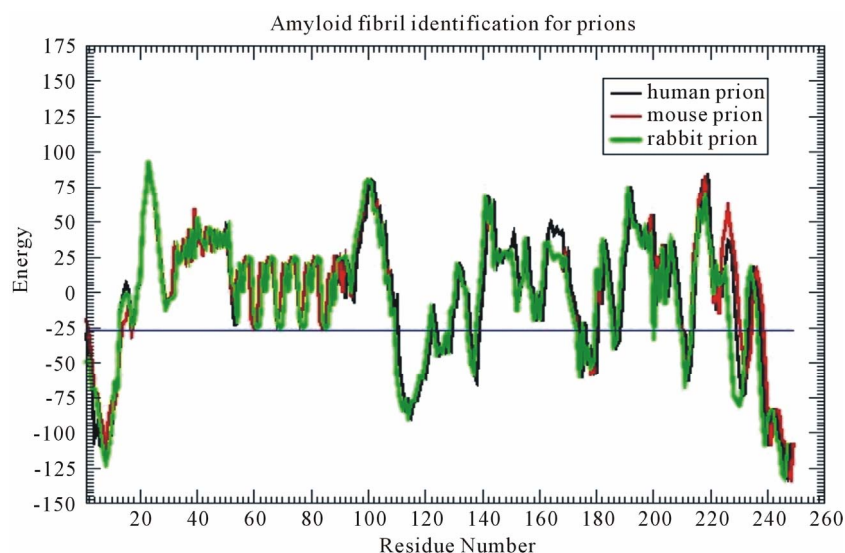


Figure 3. Prion AGAAAAGA (113 - 120) segment is clearly and surely identified as the amyloid fibril formation region, because its energy is less than the amyloid fibril formation threshold energy -26 kcal/mol.

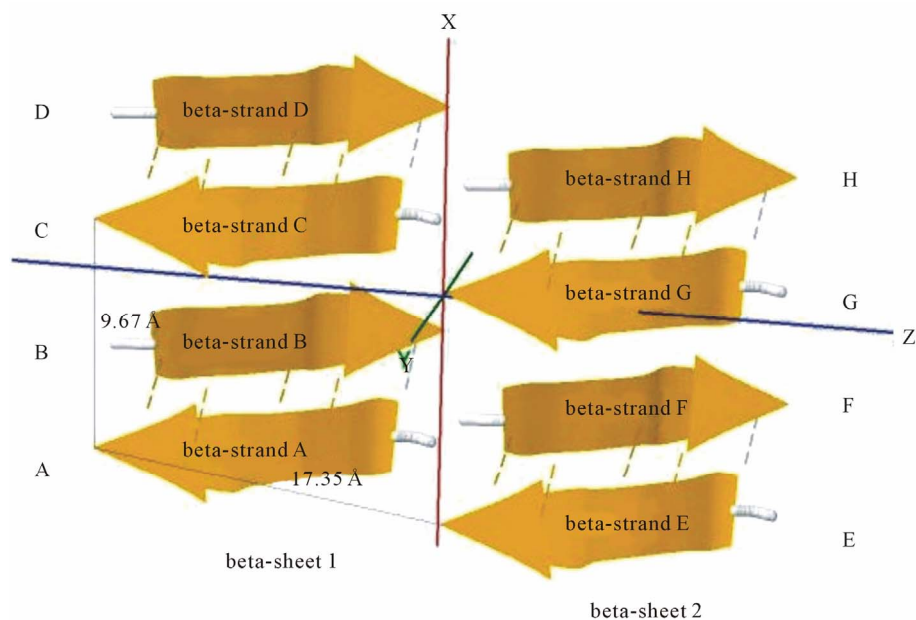


Figure 4. Protein fibril structure of human insulin LYQLEN (13 - 18) (PDB id: 2OMP). Dashed lines denote the HBs between the pairs of β -strands. A, B, C, D, E, F, G, H denote the chains of the fibril. The pair of β -sheets 1 & 2 forms a completely dry interface by vdWs, and between many pairs of β -sheets wet interfaces are formed with water molecules.

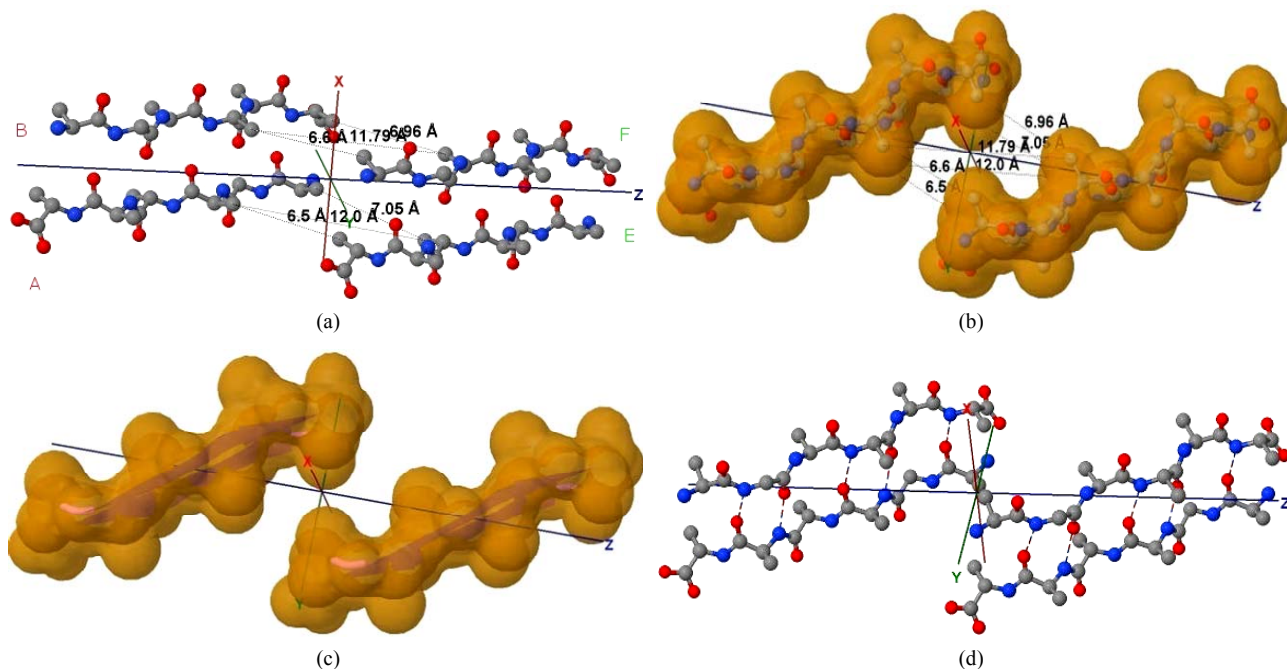


Figure 5. (a) shows the distances of “Zipper 1”-E.ALA6.CB-A.ALA3.CBE.ALA4.CB-A.ALA1.CB are 6.5, 12.0 and 7.05 angstroms respectively, and the distances of “Zipper 2”-F.ALA1.CB-B.ALA4.CB-F.ALA3.CB-B.ALA6.CB are 6.6, 11.79, 6.96 angstroms respectively. In (a), A, B, E, F denote A, B, E, F Chains, the red colored spheres are oxygen atoms, the blue colored spheres are nitrogen atoms, and the gray colored spheres are carbon atoms; (b) shows the far vdW surface; (c) shows the violet colored A, B, E, F Chains of **Figure 4**; (d) shows HBs (in turns from left to right): A/E.ALA5.O-B/F.GLY2.N, A/E.ALA5.N-B/F.GLY2.O, A/E.ALA3.O-B/F.ALA4.N, A/E.ALA3.N-B/F.ALA4.O, A/E.ALA1.O-B/F.ALA6.N.

with an initial solution (−0.067, 5.274, 7.860; −1.119, 1.311, 13.564). Similarly, in “Zipper 2”, fixing the coor-

dinates of B.ALA4.CB, B.ALA6.CB and letting the coordinates of F.ALA1.CB, F.ALA3.CB be variables, we get

another optimization problem:

$$\begin{aligned}
 & \text{minimize} \\
 & 4 \left\{ 1 / \left[\left[(x_{11} - 5.446)^2 + (x_{12} - 2.796)^2 + (x_{13} - 2.662)^2 \right]^6 \right] \right. \\
 & \quad \left. - 1 / \left[\left[(x_{11} - 5.446)^2 + (x_{12} - 2.796)^2 + (x_{13} - 2.662)^2 \right]^3 \right] \right\} \\
 & + 4 \left\{ 1 / \left[\left[(x_{21} - 5.446)^2 + (x_{22} - 2.796)^2 + (x_{23} - 2.662)^2 \right]^6 \right] \right. \\
 & \quad \left. - 1 / \left[\left[(x_{21} - 5.446)^2 + (x_{22} - 2.796)^2 + (x_{23} - 2.662)^2 \right]^3 \right] \right\} \\
 & + 4 \left\{ 1 / \left[\left[(x_{21} - 5.201)^2 + (x_{22} + 1.125)^2 + (x_{23} - 7.873)^2 \right]^6 \right] \right. \\
 & \quad \left. - 1 / \left[\left[(x_{21} - 5.201)^2 + (x_{22} + 1.125)^2 + (x_{23} - 7.873)^2 \right]^3 \right] \right\}
 \end{aligned} \tag{3.6}$$

or

$$\begin{aligned}
 & \text{minimize} \\
 & \frac{1}{2} \left\{ (x_{11} - 5.446)^2 + (x_{12} - 2.796)^2 + (x_{13} - 2.662)^2 - 3.42^2 \right\}^2 \\
 & + \frac{1}{2} \left\{ (x_{21} - 5.446)^2 + (x_{22} - 2.796)^2 + (x_{23} - 2.662)^2 - 3.42^2 \right\}^2 \\
 & + \frac{1}{2} \left\{ (x_{21} - 5.201)^2 + (x_{22} + 1.125)^2 + (x_{23} - 7.873)^2 - 3.42^2 \right\}^2 \\
 & - 0.05(x_{11} + x_{12} + x_{13} + x_{21} + x_{22} + x_{23})
 \end{aligned} \tag{3.7}$$

with an initial solution (4.714, 4.878, 8.881; 4.170, 1.360, 14.292). Next, we solve **Eqs.3.5** and **3.7** by Algorithm 1.

We first solve **Eq.3.5** in the use of Algorithm 1. We set $\sigma_1 = 10^{-4}$, $\sigma_2 = 0.1 - 0.9$, $m_1 = 3 - 7$, take the initial solution $x_0 = (-0.067, 5.274, 7.860; -1.119, 1.311, 13.564)$ and calculate its gradient $g_0 = (-69.7747, 140.135, 365.852, -752.075, -285.005, 3576.69)$ and its Hessian matrix

$$\begin{aligned}
 H_0 = & \left((66.4497, -10.415, -27.1845, 0, 0, 0)^T, \right. \\
 & (-10.415, 82.2093, 54.6557, 0, 0, 0)^T, \\
 & (-27.1845, 54.6557, 203.929, 0, 0, 0)^T, \\
 & (0, 0, 0, 380.664, -4.02618, -159.578)^T, \\
 & (0, 0, 0, -4.02618, 369.586, -25.5081)^T, \\
 & \left. (0, 0, 0, -159.578, -25.5081, 1048.02)^T \right)
 \end{aligned} \tag{3.8}$$

which is a positive definite matrix with eigenvalues (1085.02, 372.093, 341.157, 230.049, 61.2695, 61.2695). Then Algorithm 1 hybridized with simulated annealing global optimal search (in order to bring local optimal solutions to jump out of local traps, replacing the discrete

gradient local optimal search method in Algorithm 1 of [14] by the Algorithm 1 of this paper) is executed and the optimal solution (3.027, 4.954, 3.856; 1.679, 1.777, 5.011) for **Eq.3.5** is got.

Similarly, for **Eq.3.7**, we take the initial solution $x_0 = (4.714, 4.878, 8.881; 4.170, 1.360, 14.292)$ and calculate its gradient $g_0 = (-46.8782, 133.142, 397.798, -401.192, -182.604, 3436.46)$ and its Hessian matrix

$$\begin{aligned}
 H_0 = & \left((66.1163, -6.0961, -18.2092, 0, 0, 0)^T, \right. \\
 & (-6.0961, 81.3119, 51.7918, 0, 0, 0)^T, \\
 & (-18.2092, 51.7918, 218.677, 0, 0, 0)^T, \\
 & (0, 0, 0, 339.302, -2.9188, -85.8315)^T, \\
 & (0, 0, 0, -2.9188, 361.487, -2.99786)^T, \\
 & \left. (0, 0, 0, -85.8315, -2.99786, 1034.38)^T \right),
 \end{aligned} \tag{3.9}$$

which is a positive definite matrix with eigenvalues (1044.83, 361.8, 328.538, 238.159, 63.973, 63.973). The optimal solution got for **Eq.3.7** is (7.412, 4.760, 4.624; 5.887, 1.451, 5.757).

We set (3.027, 4.954, 3.856; 1.679, 1.777, 5.011) as the

coordinates of E.ALA6.CB and E.ALA4.CB, (7.412, 4.760, 4.624; 5.887, 1.451, 5.757) as the coordinates of F.ALA1.CB and F.ALA3.CB, and taking the average value we get

$$E(F) = A(B) + (1.0335, 1.0823, 0.9723). \quad (3.10)$$

By **Eq.3.10** we can get very close vdW contacts between A(B) chains and E(F) chains (**Figure 5(b)**), and other chains of Model 1 can be got by **Eqs.3.2, 3.3** and **Eq.3.11**:

$$I(J) = A(E) - (9.666, 0, 0). \quad (3.11)$$

The initial structure of Model 1 illuminated in **Figures 6(a)** and **(b)** is not the optimal structure with the lowest total potential energy. The initial structure also has no hydrogen atoms (so no hydrogen bonds existed) and water molecules added. For each Chain, the C-terminal and N-terminal atoms also have problems. Clearly there are a lot of close/bad contacts between β -strand atoms as illuminated in **Figures 6(a)** and **(b)**. We used the ff03 force field of AMBER 11, in a neutral pH environment. The amyloid fibrils were surrounded with a 8 angstroms layer of

TIP3PBOX water molecules using the XLEaP module of AMBER 11. 1944 waters and 408 hydrogen atoms were added for Model 1 by the XLEaP module. solvated amyloid fibril was minimized by the method.

The LJ potential energy of atoms' vdW interactions is just a part of the total potential energy of a protein, and by observations from Model 1 computed by **Eqs.3.2, 3.3, 3.10** and **3.11** we can see that the contacts between β -strand atoms and β -sheet atoms are too close/bad. Thus, we need to relax Model 1 computed. The relaxation is done in the use of local search LBFGS Quasi-Newton method (lbfgs_memory_depth = 3) within AMBER 11 [23]. The relaxed/optimized Model 1 is illuminated in **Figure 6(c)**. Seeing **Figure 6(d)** compared with **Figure 6(b)**, we may know the vdW interactions between the two β -sheets are very relaxed/optimized now. **Figure 6(c)** shows the Model 1 of optimal molecular structure for prion AGAAAAGA amyloid fibrils.

4. CONCLUSIONS

In a (macro) molecular system, a robust energy mini-

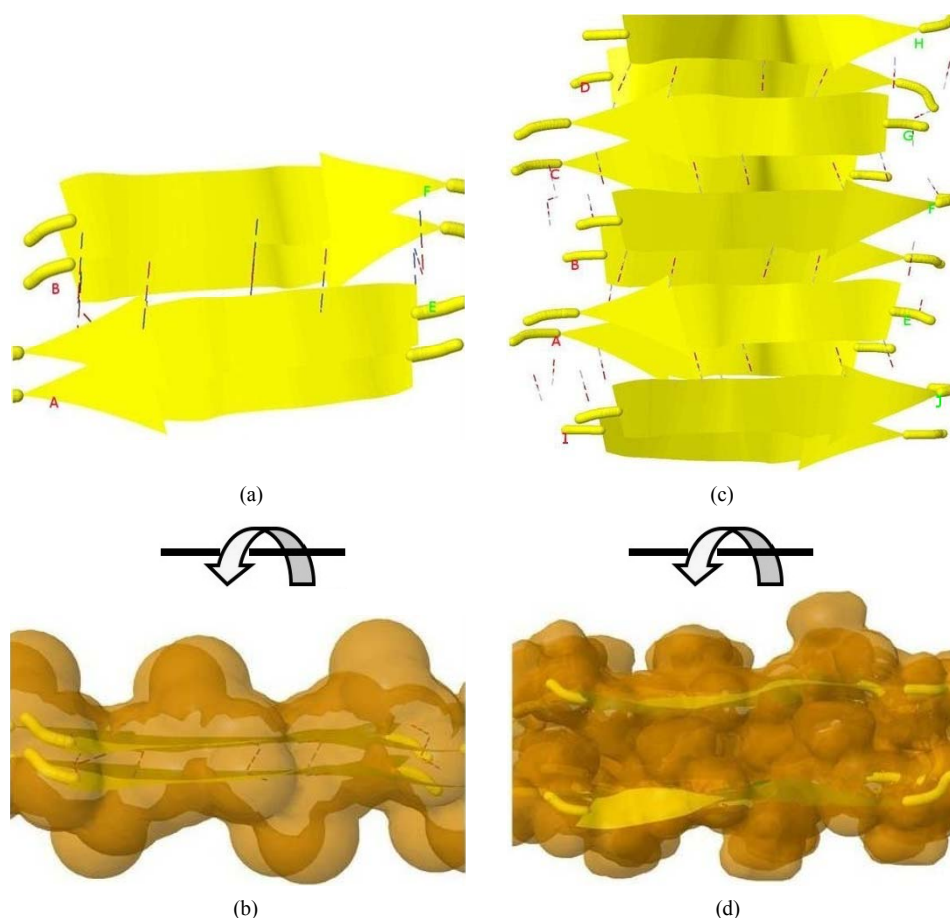


Figure 6. Left: The close vdW surface contacts between Chains A(B) and E(F) of Model 1 after solving **Eqs.3.5** and **3.7**; Right: The constructed Model 1 with 10 chains. The dashed lines denote HBs.

mization is very necessarily required. Mathematical optimization minimization methods find a place to apply in these systems. Because in physics the (macro) molecular system usually is not a simple two-body problem of system, local search optimization methods are very useful in the applications to the (macro) molecular system. On another sense, when a protein is unstable, noncrystalline or insoluble and very difficult to detect its 3D structure by the expensive and costly NMR and X-ray, theoretical mathematical or physical computational method can be used to produce the 3D structure of the protein. Moreover, even though the X-ray crystallography finds the X-ray final structure of a protein, we still need refinements using theoretical protocols in order to produce a better structure. The theoretical computational method—an improved LBFGS Quasi-Newtonian mathematical optimization method—presented in this paper and other mathematical optimization methods mentioned in this paper should be very useful in the protein molecular modeling research field.

This paper also shows the effectiveness of the improved LBFGS mathematical optimization method presented. Prion AGAAAAGA amyloid fibrils have not much structural information. This paper presents some bioinformatics on the molecular structures of prion AGAAAAGA amyloid fibrils in the sense of theoretical emphasis. The structures may be helpful in the advance in the biochemical knowledge of prion protein misfolding or instability and in the future applications for therapeutic agent design.

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